

Characterizing the Structure–Function Relationship of a Naturally Occurring RNA Thermometer

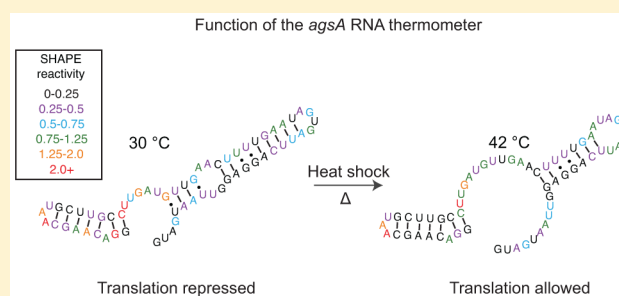
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Supporting Information

ABSTRACT: A large number of bacteria have been found to govern virulence and heat shock responses using temperature-sensing RNAs known as RNA thermometers. A prime example is the *agsA* thermometer known to regulate the production of the AgsA heat shock protein in *Salmonella enterica* using a “fourU” structural motif. Using the SHAPE-Seq RNA structure-probing method *in vivo* and *in vitro*, we found that the regulator functions by a subtle shift in equilibrium RNA structure populations that leads to a partial melting of the helix containing the ribosome binding site. We also demonstrate that binding of the ribosome to the *agsA* mRNA causes changes to the thermometer structure that appear to facilitate thermometer helix unwinding. These results demonstrate how subtle RNA structural changes can govern gene expression and illuminate the function of an important bacterial regulatory motif.



Bacterial survival depends on a bacterium’s ability to react appropriately to environmental cues, altering its behavior in response to its ever-changing surroundings. In particular, variations in temperature require coordinated systemwide responses to avoid deleterious effects,¹ resulting in the evolution of a wide array of molecular thermosensors. Among these thermosensors, RNA thermometers have been shown to control gene expression in response to changes in external temperature, allowing bacteria to govern widespread responses to heat shock and to control temperature-dependent virulence effects,² an important feature for pathogens with warm-blooded hosts.

RNA thermometers generally govern heat shock response post-transcriptionally by relying on the formation of temperature-sensitive secondary structures to fully or partially occlude the ribosome binding site (RBS) of the controlled gene,^{3,4} thus decreasing translation efficiency⁵ at low temperatures. An increase in temperature changes this structure and allows translation in a temperature-dependent manner. The most widely characterized RNA thermometers make up the broad family of ROSE (repression of heat shock gene expression) elements common among α - and γ -proteobacteria^{6,7} and found in the 5′ untranslated regions (UTRs) of heat shock genes.¹ ROSE elements share a conserved sequence motif that sequesters the Shine-Dalgarno (SD) sequence in a hairpin through an unusual base triple and noncanonical nucleotide interactions.⁸

Other heat shock RNA thermometers involve the formation of complex structures that block ribosome entry without

directly pairing with the RBS^{9,10} or depend upon an internal hairpin bulge to allow for stem melting in response to an increase in temperature.^{11,12} This hairpin bulge pattern occurs in the *Salmonella enterica agsA* RNA thermometer (Figure 1A), where an internal stem bulge is followed by a stretch of four uridines paired to the Shine-Dalgarno sequence. This structure confers thermosensing capability to the 5′ UTR of the *agsA* gene encoding aggregation suppression protein AgsA,¹³ a molecular chaperone that forms part of the heat shock response in *Salmonella*.¹⁴ Although the fourU RNA thermometer motif was first coined in reference to the *agsA* thermometer from *Salmonella*,¹³ the previously reported virulence control mechanism of the plague bacterium *Yersinia pestis* also relies on four uridines base-pairing to the Shine-Dalgarno sequence to grant temperature-dependent translational control.¹⁵ Similarly, the temperature-dependent virulence response of *Vibrio cholerae* was recently tied to a fourU element within the 5′ UTR of the *toxT* gene, a key transcriptional activator of virulence factors.¹⁶ While other temperature-based virulence controls rely on alternate mechanisms,^{17,18} the reoccurrence of the fourU motif in regulating both heat shock and virulence underlines its widespread utility as an RNA thermometer.

The fourU motif hairpin is intuitively thought to function by simply opening at elevated temperatures to allow ribosome access, though it is unclear exactly how “open” the structure must be to implement temperature-dependent control. In fact,

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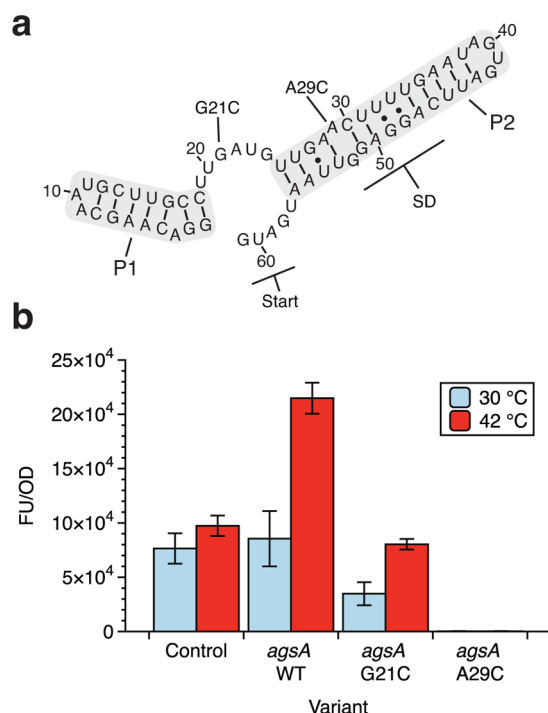


Figure 1. Structure and function of the *agsA* thermometer. (a) Putative structure of the *agsA* 5' untranslated region (UTR) RNA thermometer from *S. enterica* with key P1 and P2 helices, the Shine-Dalgarno (SD) sequence, the start codon, and mutations of interest from ref 13 labeled. (b) Functional *in vivo* testing of the *agsA* thermometer and mutants in *Escherichia coli* using the superfolder green fluorescent protein (SFGFP) reporter gene placed downstream of the 5' UTR. Cultures were grown at 30 °C for 4 h and then subjected to 30 or 42 °C for 30 min to measure the response of the thermometer constructs to heat shock. The G21C mutation extends the P2 helix to include the start codon, resulting in lower fluorescence in both the low-temperature conditions and the heat shock conditions. The A29C mutation closes the inner hairpin loop within the P2 helix, causing complete repression of gene expression under both temperature conditions. Fluorescence is normalized as fluorescence units over optical density (FU/OD).

agsA thermometer structural characterization *in vitro* via nuclear magnetic resonance (NMR) spectroscopy has provided key insights into the relative stabilities of the different pairings within the thermometer hairpin¹⁹ and the magnesium dependence of hairpin melting.²⁰ This previous work has also shown that only a few internal bases have increased flexibility at elevated temperatures, rather than large structural changes indicating melting of the entire helix.

Here we sought to improve our understanding of the mechanistic details of the *agsA* fourU thermometer by studying for the first time how *in vivo* structural changes are coupled to gene expression. In recent work, SHAPE-based probing has been deployed to explore similar RNA thermometer mechanisms *in vitro*.^{21,22} Now, by combining in-cell SHAPE-Seq to probe RNA structures *in vivo*, fluorescent reporter measurements of gene expression, and experimentally restrained computational structure prediction, we present a detailed view of the *agsA* thermometer structural ensemble and its response to heat shock. We then dissect the underlying structural mechanism through *in vitro* experiments performed in cell-free gene expression systems and highlight the effect of ribosome binding and translation on the structure of the *agsA* 5' UTR. This detailed study of the *agsA* thermometer

illuminates the intriguing relationship between subtle changes in its RNA structure and large changes in its regulatory function.

EXPERIMENTAL PROCEDURES

Cloning and Plasmid Construction. The *agsA* 5' UTR sequence from ref 13 in both its wild-type (WT) and G21C and A29C point mutant forms was cloned into a p15a plasmid backbone harboring chloramphenicol resistance. Using inverse polymerase chain reaction (iPCR) for scarless insertion, the *agsA* sequence was introduced directly between the *Escherichia coli* sigma 70 consensus promoter J23119 SpeI [a variant of the BBa_J23119 promoter from the Registry of Standard Biological Parts (http://partsregistry.org/Main_Page) in which the final six nucleotides have been replaced with a SpeI cut site] and the start codon for the superfolder green fluorescent protein (SFGFP) reporter gene. The constitutive gene expression control plasmid has a sequence designed to allow the ribosome full access to the RBS. See the Supporting Information for plasmid maps (Figure S1) and sequences (Table S1).

In Vivo Bulk Fluorescence Assays and In-Cell SHAPE-Seq. All bulk fluorescence and in-cell SHAPE-Seq experiments were performed in *E. coli* strain TG1 [*supE thi-1 Δ(lac-proAB) Δ(mcrB-hsdSM)5 (r_K⁻ m_K⁻)* [*F'* *traD36 proAB lacI^q ΔZΔM15*]] with three independent biological replicates. The overall heat shock protocol for testing the responsiveness of *agsA* to heat shock is depicted in Figure S2. For each independent replicate, an empty control plasmid (JBL 001), constitutive gene expression control plasmid (JBL2348), and *agsA* WT and mutant plasmids were transformed separately into chemically competent *E. coli* TG1 cells, plated on LB/Agar plates with 34 mg/mL chloramphenicol, and incubated overnight at 37 °C for approximately 17 h. After being incubated overnight, the plates were removed from the incubator and kept at room temperature for approximately 7 h. For each of the three replicates, three separate colonies were picked for the control and one of each tested variant, and each colony was used to inoculate 1 mL of LB medium with 34 mg/mL chloramphenicol in a 2 mL 96-well block. The block was covered with a breathable seal and incubated at 37 °C while being shaken at a speed of 1000 rpm in a benchtop shaker for 18 h overnight. From these overnight cultures, 24 μL was taken and used to inoculate two duplicate 96-well blocks containing 1.2 mL of LB medium with 34 mg/mL chloramphenicol. The blocks were shaken at 30 °C and 1000 rpm for 4 h, and then one of the blocks was shaken at 42 °C (heat shock condition) and the other kept at 30 °C for 30 min (no heat shock condition). From there, in-cell SHAPE-Seq probing (for RNA structure determination) and bulk fluorescence sample preparation (for gene expression measurement) were performed as described by Watters et al.^{23,24} using 1 mL of the exponential phase culture for in-cell SHAPE-Seq and 150 μL for the bulk fluorescence measurements. Any deviations from the published protocol are described below. The samples' optical density (OD) at 600 nm and fluorescence (485 nm excitation, 520 nm emission) were measured with a Biotek Synergy H1 plate reader for bulk fluorescence. Following RNA probing as described previously,²³ the modified and unmodified RNAs were extracted using the TRIzol Max Bacterial RNA Isolation Kit (Thermo Fisher). The extracted RNAs were reverse transcribed from within the SFGFP gene (CAACAAGAATTGGGACAACTCCAGTG primer) using 3 μL of 50 nM primer per reaction. The following RNA hydrolysis step was extended from 5 to 15 min, and the

partial neutralization with HCl was omitted. Subsequent ethanol precipitation, adaptor ligation, and bead purification steps followed the published protocol exactly,²³ and quality analysis and library construction were performed following the previously reported PCR modifications to minimize side-product formation for libraries from weakly expressed RNAs.²³ Data analysis was executed using Spats pipeline version 1.0.1 as described by Watters et al.²⁴ to generate raw θ reactivities,²⁵ which were then converted to ρ reactivities by multiplying by the length of the probed RNA²⁶ and normalized using the box-plot strategy described by Low and Weeks.²⁷ The normalization method ignores outlier reactivities, defined as those reactivities more than 1.5 times the interquartile range (IQR) above the 75th percentile, and then normalizes all reactivities by the average of the next 10% of reactivities. This normalization scheme is designed to allow for comparison of reactivities between structures with varying distributions of paired and unpaired nucleotides.

mRNA Preparation. DNA templates for *in vitro* transcription were generated using PCR to add the T7 promoter to the *agsA* variant or control sequences and to amplify the region from the 5' UTR to the intrinsic terminator following the SFGFP gene. The resulting templates were transcribed *in vitro* with T7 polymerase, and the resulting mRNAs were purified using Ampure XP beads (Beckman Coulter) following the manufacturer's protocol and run alongside an RNA ladder on a 3% denaturing polyacrylamide gel. The gel was stained using SYBR Gold (Thermo Fisher) and visualized under ultraviolet light to check for template length and purity.

PURExpress Assays. The cell-free assays showing temperature-sensitive expression from both *agsA* DNA templates and pretranscribed mRNA were performed in the PURExpress Δ Ribosome Kit (New England Biolabs). To each transcription-translation reaction mixture were added 4 μ L of kit solution A, 1.2 μ L of factor mix, 1.8 μ L of *E. coli* ribosomes (New England Biolabs), 0.75 μ L of *E. coli* RNA polymerase holoenzyme (New England Biolabs), 0.4 μ L of SUPERase In RNase Inhibitor (Thermo Fisher), 100 ng of plasmid template, and enough nuclease-free water to reach a total volume of 10 μ L. Each translation-only reaction mixture contained 4 μ L of solution A, 1.2 μ L of factor mix, 1.8 μ L of *E. coli* ribosomes, 3.24 pmol of purified mRNA template, and enough nuclease-free water to reach a total volume of 10 μ L. Reaction mixtures were pipetted into a clear-bottom, black 384-well plate loaded onto a Biotek Synergy H1 plate reader prewarmed to either 30 or 42 °C, where fluorescence (485 nm excitation, 520 nm emission) was monitored for 3 h with measurements taken every 5 min. Measurements were corrected for background fluorescence by subtracting the average reading for three empty wells.

In Vitro SHAPE-Seq in Folding Buffer. *In vitro* SHAPE-Seq was performed largely following the *in vitro* SHAPE-Seq protocol of Watters et al.²⁴ performed on 10 pmol of purified mRNA folded and probed at 30, 42, or 80 °C in 18 μ L of folding buffer (final concentrations of 10 mM MgCl₂, 100 mM NaCl, and 100 mM HEPES). The folded RNA was split into two 9 μ L aliquots and probed with the addition of 1 μ L of 65 mM 1M7 (1-methyl-7-nitroisatoic anhydride) in dimethyl sulfoxide (DMSO) (positive channel) or 1 μ L of DMSO (negative channel). Folding and modification steps were altered slightly in experiments comparing 30 and 80 °C to match the conditions of Wilkinson et al.²⁸ Specifically, RNAs were equilibrated at 30 °C for 20 min before being heated to the

final folding temperature (30 or 80 °C) for 5 min. Additionally, 1M7 and DMSO were prewarmed for 2 min before modification. After modification, the RNAs were precipitated with ethanol with the addition of 90 μ L of nuclease-free water, 10 μ L of 3 M sodium acetate (pH 5.5), 1 μ L of glycogen, and 300 μ L of ice-cold 100% ethanol, followed by freezing for 30 min at −80 °C and centrifugation at 4 °C and 15000 rpm for 30 min. All liquid was aspirated, and the pellets were washed with 200 μ L of 70% ethanol before further centrifugation for 2 min, removal of the wash liquid, and resuspension in 10 μ L of nuclease-free water. From there, the remainder of the experimental protocol, from reverse transcription through raw reactivity calculation, followed the published method of Watters et al.²⁴ Reactivities were normalized as described above.

In Vitro SHAPE-Seq in PURExpress. For experiments that probed the structure of the *agsA* WT and control mRNAs in PURExpress without ribosomes, with active translation, or with stalled translation initiation complexes, 3.24 pmol of purified mRNA was probed at either 30 or 42 °C. The translation initiation complex reaction mixtures were assembled using the PURExpress Δ aa Δ tRNA kit (New England Biolabs), with each 10 μ L reaction mixture containing 2 μ L of kit solution A, 1 μ L of 3 mM L-methionine, 1 μ L of kit tRNA, 3 μ L of kit solution B, and 3.24 pmol of mRNA; the lack of amino acids other than methionine was designed to cause ribosomal complex stalling at the start codon. The PURExpress probing experiments without ribosomes were performed in a similar fashion using the PURExpress Δ Ribosome Kit (New England Biolabs), with each 10 μ L reaction mixture containing 4 μ L of kit solution A, 1.2 μ L of factor mix, and 3.24 pmol of mRNA. The PURExpress active translation reaction mixtures were assembled identically except for the addition of 1.2 μ L of *E. coli* ribosomes. PURExpress reaction mixtures were incubated for 20 min (no ribosomes and stalled translation initiation complexes) or 35 min (active translation) at 30 or 42 °C before being split for modification; 5 μ L of the reaction mixture was added to 0.56 μ L of 65 mM 1M7 in DMSO (positive channel) and to 0.56 μ L of DMSO (negative channel) and incubated for an additional 1 min before being transferred to ice. From there, the modified RNA and unmodified RNA were extracted using the Trizol Max Bacterial RNA Isolation Kit (Thermo Fisher), and the remainder of the protocol (including reactivity normalization) followed the same method as the in-cell SHAPE-Seq samples.

Statistical Analysis and Data Presentation. Experiments were performed with three distinct replicates. Error bars represent the sample standard deviation, and significance was determined using two-sided heteroscedastic Welch's *t* tests. For SHAPE-Seq difference plots, the error was propagated using standard propagation of error techniques. Reactivity normalization was performed as described in the in-cell SHAPE-Seq section above.

RNAstructure Structure Prediction and MDS Analysis. Minimum free energy (MFE) RNA structures were predicted using the Fold command from the RNAstructure command-line interface.²⁹ Box-plot normalized reactivities were specified as pseudoenergy restraints using RNAstructure's "sh" option, and the SHAPE restraint intercept (*b*) and slope (*m*) were set at −0.3 and 1.1 kcal/mol, respectively, on the basis of the optimum values for weighting ρ reactivities as determined by Loughrey et al.²⁶ Temperature was specified as either 303.15 K (30 °C) or 315.15 K (42 °C) using the "t" temperature option. Structures were output as connectivity (ct) files, converted to dot-bracket notation (dbn) using the ct2dot utility from

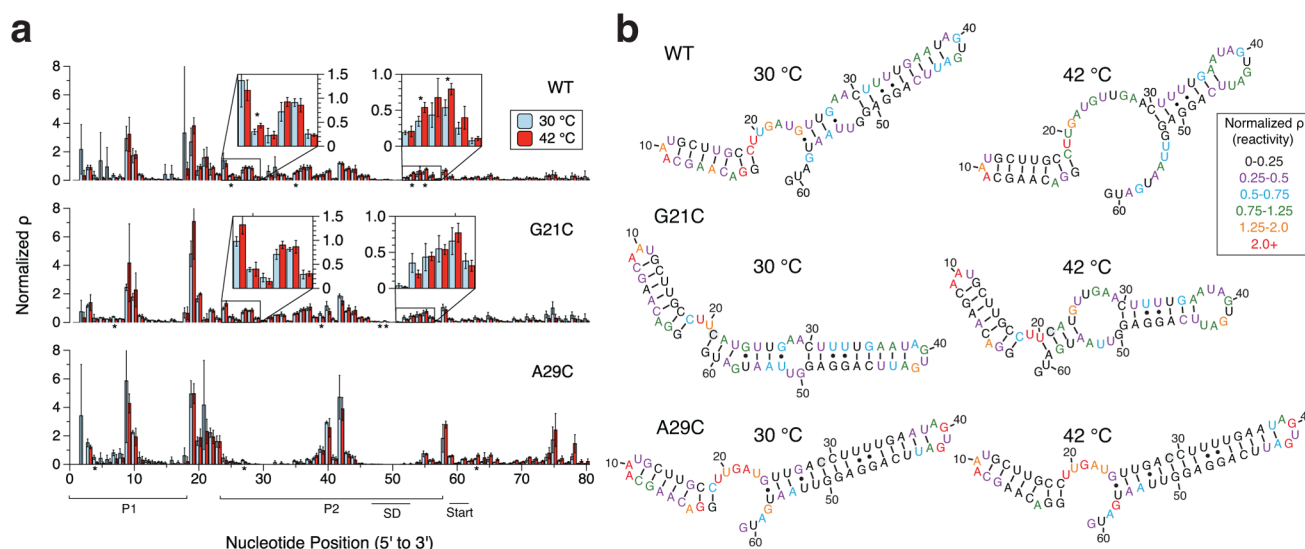


Figure 2. SHAPE-Seq characterization of the *agsA* thermometer *in vivo*. (a) SHAPE-Seq normalized reactivity (ρ) plots for *agsA* WT, G21C, and A29C variants from triplicate experiments *in vivo* performed at 30 °C and using 42 °C heat shock conditions. Error bars represent standard deviations over replicates, and stars represent statistical significance between 30 and 42 °C reactivities. (b) Experimentally informed minimum free energy (MFE) RNA structures as predicted using the RNAstructure Fold program with the reactivity values from panel a taken as pseudoenergy folding restraints.²⁹ Nucleotides are color-coded according to normalized SHAPE reactivity (ρ), with red indicating high reactivity and black low reactivity. Point-mutated nucleotides are shaded in gray.

RNAstructure, and drawn using Adobe Illustrator (Figures 2b, 3d, 5b, S7, and S10b) or the VARNA graphical user interface (Figures S3–S6).³⁰ Structures in Figures 2b, 3d, 5b, S7, and S10b represent predictions for *agsA* variant 5' UTRs. Structures including the full mRNA sequence restrained by SHAPE data for the 5' UTR and SFGFP leader sequence are presented in Figures S3–S6. Partition function generation for the RNA-probability-pairing matrices was performed using the RNAstructure web server's partition function, with the same temperature and pseudoenergy SHAPE restraints as the MFE structure prediction, and probability plots were generated using the ProbabilityPlot function. For Figure S8, the desktop version of RNAstructure was used to generate the text-based ProbabilityPlot using the -t option. For multidimensional scaling (MDS) analysis of the RNA structural ensemble, 100000 structures were sampled from each condition using the RNAstructure stochastic function. Following ref 31, all unique structures were coded as binary vectors based on whether each nucleotide was base-paired (0, not base-paired; 1, base-paired), and the resulting binary vectors were subjected to two-component multidimensional scaling performed with the R Project for Statistical Computing (version 3.2.3).

RESULTS

Characterization of *in Vivo* Structure and Function of the *agsA* Thermometer. As a first step, we characterized the function and structure of the *agsA* thermometer *in vivo*. To measure the thermometer's temperature sensitivity, we fused the *agsA* 5' UTR directly to the superfolder green fluorescent protein (SFGFP) coding sequence and tested the construct's heat responsiveness *in vivo* in *E. coli* (Figure 1). Alongside the wild-type (WT) *agsA* thermometer, we also tested a control 5' UTR known to allow downstream translation at all temperatures and a pair of *agsA* mutants known to exhibit altered temperature responsiveness.¹³ Specifically, the G21C mutation is predicted to extend the *agsA* P2 helix to include the AUG start codon at its base, resulting in repression of translation at

30 °C that is stronger than that of the wild-type. The A29C mutation removes a predicted single base pair internal hairpin loop, thus strengthening the P2 helix and preventing RBS access at both 30 and 42 °C (Figure 1a).

To test the responsiveness of the constructs to heat shock, we transformed them into *E. coli* and grew triplicate colonies overnight in liquid media. Duplicate subcultures of each colony were grown at 30 °C for 4 h, and then one was subjected to a 30 min heat shock at 42 °C while the other remained at 30 °C (Figure S2). From these cultures, we measured both the fluorescent gene expression and the structure of the *agsA* RNAs with in-cell SHAPE-Seq.²³

Gene expression was measured by determining fluorescence and optical density of the cultures using an optical plate reader, which were used to calculate the normalized fluorescence units over optical density (FU/OD) values presented in Figure 1b. The control construct showed a slight increase in normalized fluorescence under heat shock, possibly corresponding to the faster production and maturation of SFGFP at the higher temperature. However, compared to the control, the WT *agsA* variant showed clear activation in response to heat shock, with normalized fluorescence 2.5 times that seen under the 30 °C condition. In contrast, the G21C *agsA* variant gave lower overall levels of fluorescence but still showed an increased level of gene expression in response to heat shock, while the A29C variant SFGFP expression levels were barely distinguishable above background at 30 and 42 °C (Figure 1b). These results confirm that the behavior of the *agsA* thermometer and its designed mutants in our expression configuration matches their behavior as originally reported by Walminghaus et al. using a different reporter system.¹³

Because mutations that stabilized the hairpin resulted in a decreased level of gene expression, the results described above support the model of thermometer regulation tied to hairpin melting at higher temperatures, though they do not detail the specific RNA structural changes that take place upon heat shock. To determine these structural changes *in vivo*, we probed

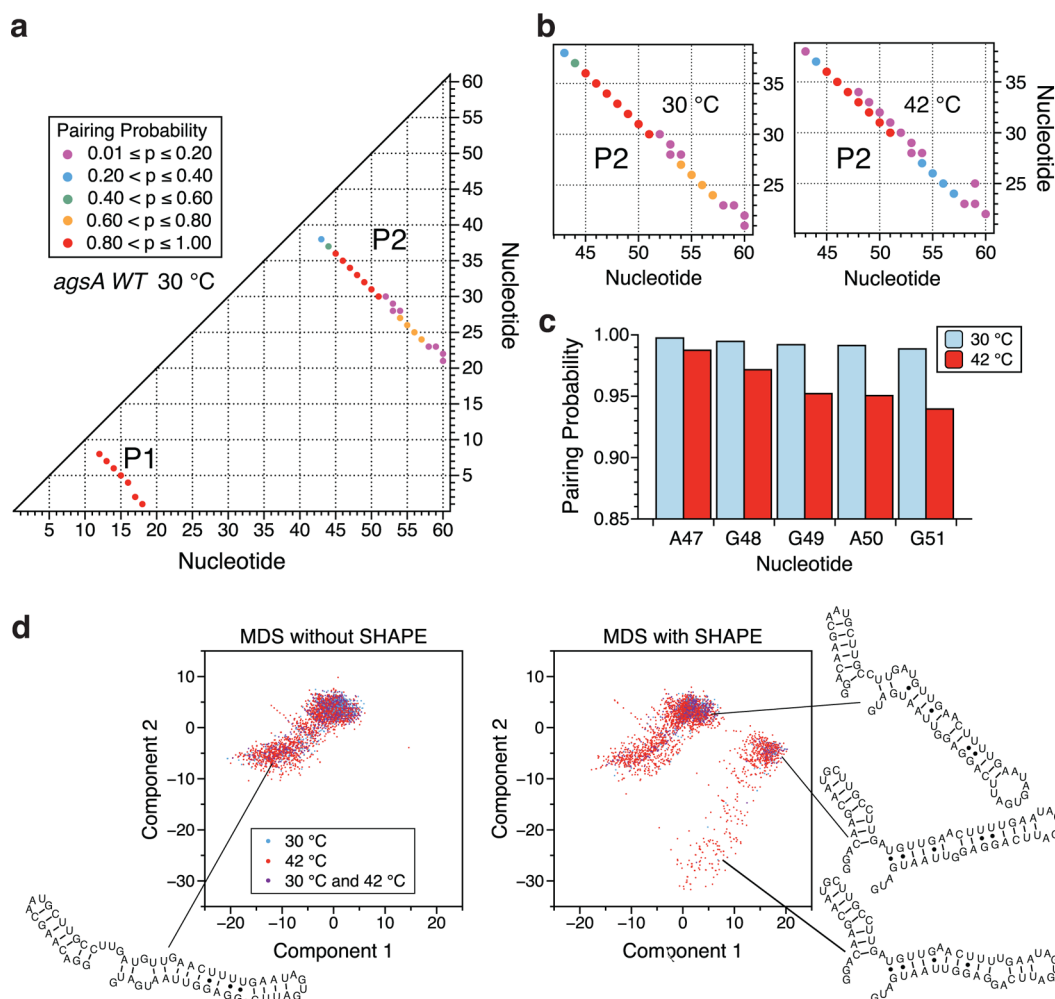


Figure 3. Ensemble visualization of the *agsA* WT thermometer-modeled secondary structures at 30 and 42 °C. (a) Pairing-probability plot for the *agsA* WT thermometer at 30 °C. Dots correspond to base pairs that occur in $\geq 1\%$ of the predicted structural ensemble, as restrained by *in vivo* SHAPE-Seq data. Nucleotide numbers are plotted along the axes, with the x and y coordinates of a dot indicating the bases involved in the pair. Dot color corresponds to pairing probability, with red being the most likely and purple the least likely. (b) Close-up of *in vivo* SHAPE-Seq-restrained pairing-probability plot for *agsA* WT thermometer hairpin 2 at 30 and 42 °C. Changes in pairing probability are indicative of the opening of the helix with an increase in temperature. (c) Base-pairing probabilities for the first five nucleotides of the Shine-Dalgarno sequence as determined by the structural ensemble predictions restrained by *in vivo* SHAPE-Seq data, plotted at both 30 and 42 °C. (d) Multidimensional scaling (MDS) representations of the structural ensemble of the *agsA* WT thermometer at 30 and 42 °C, with and without the *in vivo* SHAPE-Seq restraint of structural prediction. For each condition, 100000 structures were sampled from the corresponding partition function and visualized in two-dimensional space via MDS as described in [Experimental Procedures](#). Representative structures are shown.

the structure of the *agsA* 5' UTR using in-cell SHAPE-Seq,^{23,24,32,33} in the same cell cultures used to make gene expression measurements. SHAPE-Seq is an RNA structure probing technique that uses a small chemical probe that covalently modifies RNA nucleotides in a structure-dependent fashion.²⁵ The probe used here, 1M7 (1-methyl-7-nitroisatoic anhydride), also allows for in-cell probing of RNA structure.^{23,24,32,33} In general, members of the SHAPE family of probes preferentially react with unconstrained nucleotides like those in loops and single-stranded regions but do not frequently modify nucleotides constrained by base pairing, stacking interactions, or ligand interactions.³⁴ The locations of modifications are detected through reverse transcription and sequencing, allowing for the determination of the relative reactivities of the underlying RNA nucleotides to the SHAPE-Seq probe. High reactivities correspond to unpaired or unconstrained nucleotides, while low reactivities indicate base pairing or constrained stacking interactions. These experimen-

tally measured SHAPE-Seq reactivities can be incorporated into structural prediction programs as a pseudoenergy restraint that favors structures matching the observed reactivity patterns to arrive at highly accurate predictions of RNA secondary structure.^{26,27,29}

We applied in-cell SHAPE-Seq to the *agsA* thermometer variants using reverse transcription primers that bound to the downstream coding sequence of SFGFP. Cultures were probed at 30 and 42 °C simultaneously with the bulk fluorescence assays presented in [Figure 1](#). The resulting reactivity profiles ([Figure 2a](#)) were used to restrain RNA structural prediction ([Figure 2b](#)) using the RNAstructure software package²⁹ to predict the minimum free energy (MFE) thermometer structures at each temperature. A comparison of the reactivity profiles and MFE folds between *agsA* variants and temperatures reveals a number of interesting differences. For the wild-type *agsA* RNA, slight reactivity increases in the nucleotides comprising the base of the P2 helix under heat shock

correspond to a predicted partial melting of the P2 hairpin at 42 °C. Though the reactivity changes are small, they correlate with the functional data showing an increased level of gene expression at the higher temperature, potentially mediated by increased access of the ribosome to the Shine-Dalgarno region as the hairpin partially melts. While the G21C mutation restrained model does extend the P2 helix to contain the start codon at 30 °C, slight reactivity increases at 42 °C match a predicted expansion of the internal hairpin loop, correlating with the observed increase in fluorescence under heat shock conditions. Notably, the wild-type temperature transition was not correctly predicted by RNAstructure in the absence of SHAPE-Seq restraints (Figure S7); rather, the subtle reactivity changes measured by SHAPE-Seq influenced the folding algorithm enough to alter the predicted structures for the wild-type variant. In comparison, the A29C mutation that closes the inner hairpin loop creates a variant with strikingly low reactivities throughout the P2 helix and displays no predicted structural change between 30 and 42 °C, retaining an extended hairpin form even under heat shock. This lack of hairpin opening corresponds to nearly undetectable levels of gene expression at both temperatures, bolstering the proposed mechanism of translational control through hairpin opening and ribosome access.

Despite the overall agreement with experimentally informed predicted structural changes, it is worth noting that even at 42 °C, the MFE wild-type structure is still predicted to have near perfect base pairing of the Shine-Dalgarno sequence due to the subtle nature of the observed reactivity changes. However, the low reactivities observed in the Shine-Dalgarno sequence at 42 °C do not reflect a technical limitation of the SHAPE-Seq probing technique, as subsequent *in vitro* probing at 80 °C demonstrated clear modification of the Shine-Dalgarno nucleotides when the thermometer was further destabilized at a higher temperature (Figure S8). Thus, the slight changes observed between 30 and 42 °C suggest a model of thermometer function that is more nuanced than widespread structural changes from hairpin melting increasing access of the ribosome.¹³ Instead, the SHAPE-Seq data seem to indicate that only a small fraction of the RNA population is undergoing considerable structural changes. To examine this further, we used the RNAstructure ProbabilityPlot function to calculate the experimentally restrained ensemble base-pairing probabilities for the WT variant at 30 and 42 °C (Figure 3a,b). Points in these pairing-probability matrices reflect the probability that two bases will pair across the entire ensemble of structures, with helix elements showing up as diagonals in the matrix. Careful inspection of the probability matrices shows shifts toward lower-probability base pairing in the P2 stem at the higher temperature (Figure 3b), with Shine-Dalgarno base-pairing probabilities decreasing slightly (Figure 3c).

To obtain a more intuitive view of the population shift effect, we sampled structures from the experimentally restrained equilibrium partition function with RNAstructure's stochastic method and projected these structures onto a two-dimensional plane with multidimensional scaling (MDS) (Figure 3d). This analysis reveals that at 30 °C, the ensemble is dominated by a single cluster of similar closed structures, while at 42 °C, a small fraction of the population shifts toward structures that have some bases unpaired, but only when experimental probing data are taken into account.

Consistent with these analyses, we observe only slight changes in the reactivity of the Shine-Dalgarno sequence

(Figure 2a), with the experimentally restrained calculated ensemble-pairing probabilities shifting by just a few percentage points from approximately 99% pairing to 94–98% pairing at higher temperatures (Figure 3c). This subtle shift could correspond to a small population of transiently “breathing” hairpins. Intriguingly, this small ensemble shift in RBS accessibility is clearly enough to more than double the level of expression of the downstream reporter gene, demonstrating how subtle shifts in RNA structural ensembles can lead to large changes in functional output.

Establishing the Mechanism of the *agsA* Thermometer's Functional Response to Temperature. Having observed *agsA* thermometer structure and function within the complex cellular environment where transcription and translation happen simultaneously, we next sought to use *in vitro* studies to decouple the two processes and better discern the underlying mechanism behind the *agsA* thermometer's functional response to temperature. In particular, there is a possibility that temperature could influence the way the RNA molecule folds co-transcriptionally,³⁵ which could then impact downstream translation of the mRNA. On the other hand, a simpler and purely post-transcriptional mechanism would rely on the ensemble of existing RNA folds adjusting to a temperature change, thus leading to changes in translation.

To distinguish between the two, we first tested the latter case by performing functional gene expression assays in the *in vitro* PURExpress system from purified *agsA* mRNAs. PURExpress is an *in vitro* protein synthesis system that can be used to express proteins from plasmid DNA or from purified mRNAs.³⁶ We first established that the *agsA* thermometer variants gave the expected reporter output in PURExpress from plasmid input, allowing for both transcription and translation (Figure S9). Next, we investigated whether we could observe the same functional response to temperature from purified *agsA* mRNAs. To do so, we tracked fluorescent reporter gene expression via a plate reader for PURExpress reaction mixtures containing equal amounts of pretranscribed, purified mRNAs for the control and *agsA* variants, thus eliminating any effect of co-transcriptional folding at a particular temperature. The resulting hour-long fluorescence traces (Figure 4a) recapitulated the function observed *in vivo*, demonstrating fluorescence that is higher relative to the control at 42 °C than at 30 °C for the WT and G21C mutant and consistently low fluorescence for the A29C variant. The baseline increase in the level of gene expression of the control from 30 to 42 °C is larger than that seen *in vivo*, which could be due partly to the longer duration of the heat shock and partly to the magnifying effect of the high-yield PURExpress system. Figure 4b presents the measured rates of fluorescence increase during linear growth, reflecting the same trends observed in overall fluorescence. While these experiments cannot rule out co-transcriptional effects completely, the significant temperature responsiveness of the pretranscribed *agsA* construct mRNAs demonstrates that this RNA thermometer can function through thermodynamic shifts in the RNA folding ensemble as was previously suggested by Waldminghaus et al.¹³

Investigating *agsA* Thermometer Folding *in Vitro* Using SHAPE-Seq. Having confirmed the basic mechanism of *agsA* thermometer function, we set about investigating *agsA* thermometer folding *in vitro* to explore whether we could detect a stronger signal for structural switching in the absence of other cellular factors. Performing SHAPE-Seq *in vitro* in the PURExpress cell-free system without ribosomes on *agsA* WT

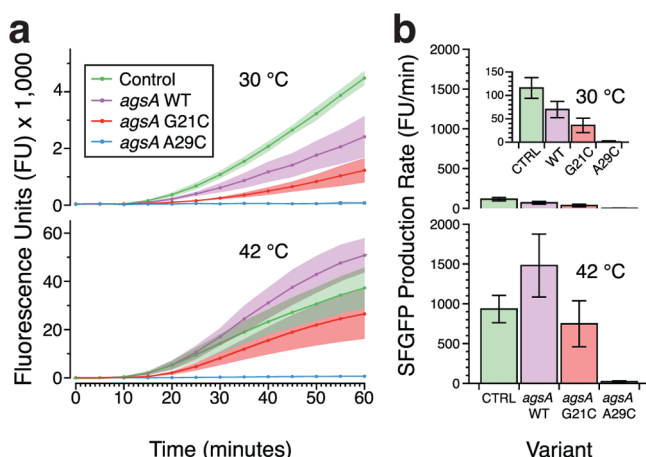


Figure 4. Testing *agsA* thermometer function in a cell-free protein synthesis system using purified pretranscribed mRNA. (a) Fluorescence trajectories over time for translation of SFGFP from *agsA* constructs and control mRNA in the PURExpress protein synthesis system at 30 and 42 °C. Shading represents the standard deviation over three replicates. (b) SFGFP production rates, calculated from the trajectories in panel a, during the linear synthesis regime for *agsA* constructs and control at 30 °C (45–50 min) and 42 °C (30–35 min), with error bars representing the standard deviation.

mRNA at 30 and 42 °C produced the reactivity plots presented in Figure 5a. Quite notably, the reactivity profiles at both temperatures are markedly similar to those measured *in vivo*, with only a handful of nucleotides showing statistically significant differences in reactivity. When the *in vitro* reactivities are used to restrain the RNA structural prediction, the resulting structural predictions (Figure 5b) parallel those seen under *in vivo* conditions, but with two key differences: at 30 °C, the internal loop in the P2 helix is predicted to be 1 bp smaller *in vitro*, and at 42 °C, the P2 stem–loop structure is predicted to be 2 bp larger *in vitro*. Similar patterns of reactivity and structure were seen for *in vitro* experiments conducted in buffer as opposed to PURExpress, demonstrating a stable pattern of

helix formation with slight variations in loop size (see Figures S10 and S11). Overall, the close agreement between *in vivo* reactivities and *in vitro* reactivities in the absence of active translation suggests that the cellular translation machinery is exploiting subtle changes in RNA helix opening to enact large changes in gene expression.

Determining the Effect of Translation and Ribosome Binding on *agsA* RNA Structure. We next sought to uncover the influence of ribosome binding on *agsA* thermometer structure. Specifically, we performed additional SHAPE-Seq experiments *in vitro* in PURExpress involving both active translation and stalled translation initiation complexes (TIC) to determine the changes in reactivity caused by ribosome binding and active translation in the clearly defined *in vitro* system (Figure 6). For the active translation experiments, we probed PURExpress reactions under the same conditions that were used for functional studies. The reactivities in the *agsA* 5' UTR are relatively similar in the no ribosome and active ribosome conditions (Figure 6b), with slight reactivity increases in the P2 helix during active translation. Interestingly, we also observed reactivity differences within the coding sequence, where reactivities are lower with active ribosomes, most likely due to actively translating ribosomes providing protection from SHAPE modification.

To probe stalled TIC complexes, we probed PURExpress reaction mixtures without all amino acids except for methionine, thus causing the translation initiation complex to stall after the addition of the first amino acid. As in the active ribosome experiment, there is a decrease in reactivity directly downstream from the ribosome binding site, likely corresponding to the ribosome footprint blocking SHAPE modification (Figure 6c). The large size of the region of the decrease in reactivity extending ~20 nucleotides beyond the start site is consistent with previous observations of stalled ribosomes in the presence of tRNA(fMet).³⁷ Even more interestingly, many of the nucleotides that formed the P2 helix, on the 5' side and the upper portion of the 3' side, undergo large increases in reactivity under the stalled TIC complex condition. This

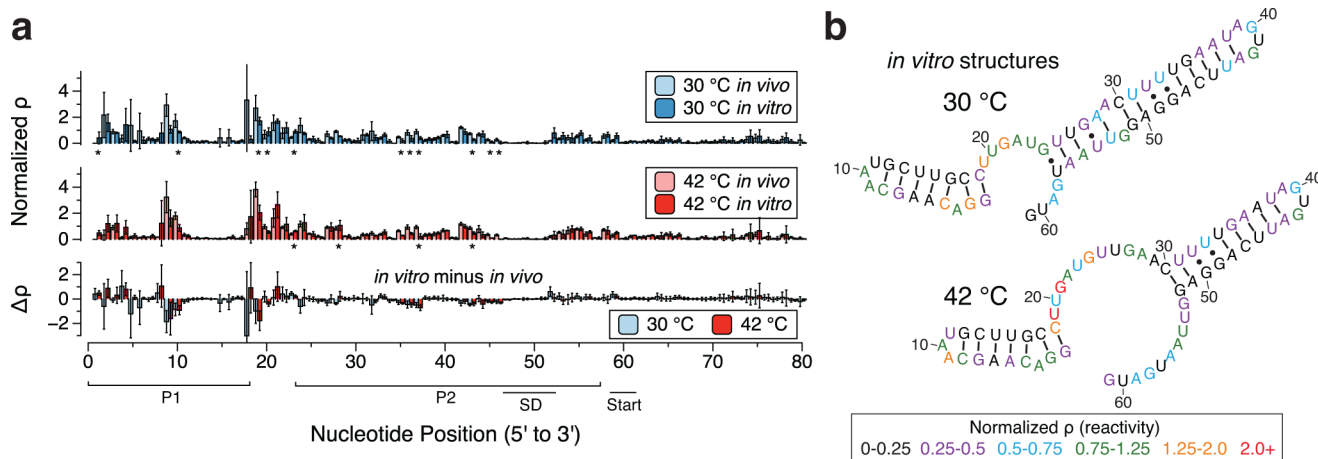


Figure 5. SHAPE-Seq characterization of the *agsA* thermometer *in vitro* in PURExpress. (a) SHAPE-Seq normalized reactivity plots for *agsA* WT mRNA from triplicate experiments *in vitro* performed on mRNAs at 30 and 42 °C, graphed alongside *in vivo* reactivities from Figure 2a for comparison. Error bars represent standard deviations over replicate measurements. Stars denote statistically significant differences between *in vivo* and *in vitro* methods as determined by heteroscedastic Welch's *t* tests, while the difference plot shows the magnitude of reactivity change (*in vitro* reactivities minus *in vivo* reactivities). (b) Experimentally informed minimum free energy (MFE) RNA structures as predicted using the RNAstructure Fold program with the *in vitro* reactivity values from panel a taken as pseudoenergy folding restraints. Nucleotides are color-coded according to normalized SHAPE reactivity (ρ), with red indicating high reactivity and black low reactivity.

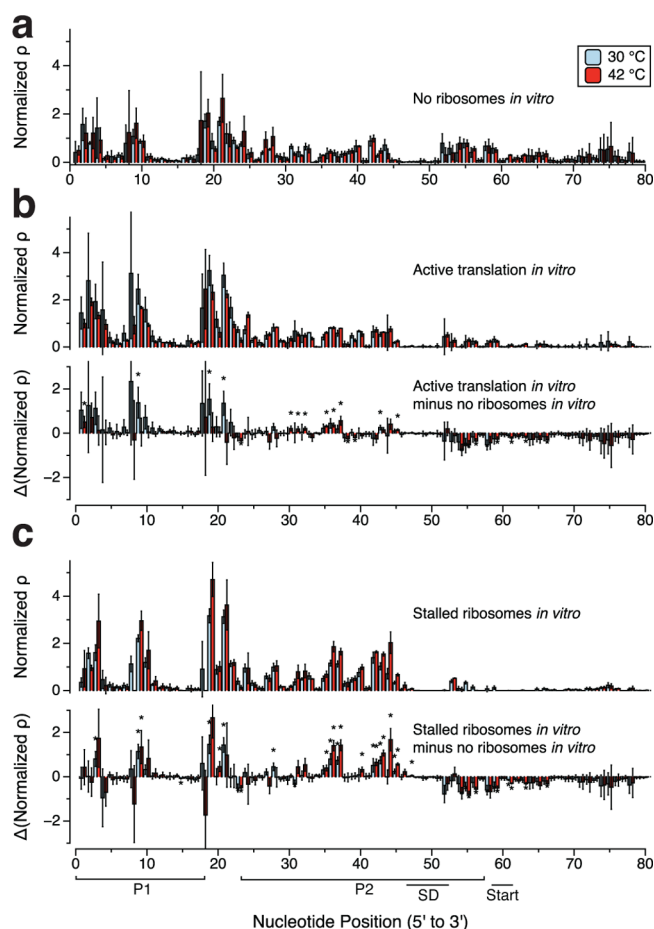


Figure 6. Investigating the effect of translation and ribosome occupancy on *agsA* WT structure and reactivity. (a) *agsA* WT SHAPE-Seq reactivity plot for triplicate experiments performed *in vitro* in PURExpress without ribosomes. (b) *agsA* WT SHAPE-Seq reactivity plot in PURExpress with active translation and difference plot of reactivities showing the change from the no ribosome case in panel a. (c) *agsA* WT SHAPE-Seq reactivity plot in PURExpress with stalled translation initiation complexes generated by supplying only methionine and difference plot of reactivities showing the change from the no ribosome case in panel a. Error bars represent standard deviations over replicate measurements. Stars represent statistical significance as in Figure 5.

suggests increased nucleotide flexibility caused by the dissolution of the P2 helix in concert with ribosome binding. Larger changes in reactivity at 42 °C than at 30 °C suggest greater ribosome access at the higher temperature, matching the functional data seen *in vivo* and *in vitro*. However, given that the ribosomes were supplied in excess of the mRNA (8:1 ratio) and allowed 20 min to reach binding equilibrium, it is unsurprising that the reactivities indicate ribosome binding to a large portion of the mRNA ensemble under both temperature conditions. The presence of background gene expression at 30 °C in functional testing indicates some level of ribosome access even at the lower temperature, potentially linked to a small fraction of the ensemble undergoing transient hairpin breathing that allows for ribosome capture. Overall, these data suggest an active role for ribosome binding in mediating additional RNA structural changes that facilitate *agsA* thermometer function.

DISCUSSION

In this work, we have combined *in vivo* and *in vitro* structure–function probing experiments to uncover detailed mechanistic aspects of how the *agsA* RNA thermometer activates gene expression in response to heat shock. Specifically, these experiments represent the first *in vivo* RNA structure probing of an RNA thermometer and nicely complement previous *in vitro* structural studies of fourU thermometers.^{13,19–21}

Overall, our *in vivo* SHAPE-Seq data revealed only small structural changes in the *agsA* thermometer with an increase in temperature, indicating subtle shifts in the underlying RNA structural ensemble upon shifting to higher temperatures. These subtle changes represent an interesting contrast to the recent *in vitro* SHAPE study of the *S. enterica htrA* fourU thermometer, which found much more significant reactivity changes between probing at 30 and 42 °C.²¹ There are two important technical differences between the study presented here and that of Edric et al. that could account for these differences in the magnitude of temperature-mediated SHAPE reactivity changes observed. The first is divalent ion concentration. Our use of 10 mM MgCl₂ under *in vitro* folding conditions rather than 2 mM MgCl₂, which was used by Edric et al., may promote the formation of stronger RNA structures that are more resistant to melting in our studies.³⁸ The second is the choice of SHAPE reagent. Interestingly, while Edric et al. used both NMIA and 1M7, they observed larger changes in reactivity with NMIA, suggesting there may be a difference between the effects of these two reagents when probing temperature-mediated structural transitions in RNAs. Despite these technical differences, there is an additional structural reason that could account for these differences: the *htrA* thermometer has the fourU-SD motif at the base of the thermometer hairpin, while the *agsA* thermometer has this motif in the upper portion of a more extended stem. The presence of the motif at the base of the hairpin, which we predict on the basis of SHAPE data to melt even in the *agsA* thermometer, could facilitate easier melting in the *htrA* context.

Our observation of minimal structural changes in the *agsA* thermometer is consistent with previous *in vitro* NMR structural studies that showed gradual base pair dissociation at higher temperatures.¹⁹ Thus, it appears that temperature shifts do not cause large-scale structural rearrangement in this simple hairpin. However, our subsequent *in vitro* functional experiments showed that these subtle equilibrium-based post-transcriptional changes in RNA structure do lead to temperature-dependent control of gene expression. This is despite the lack of major temperature-dependent structural differences in the Shine-Dalgarno region, previously proposed as the mechanism behind *agsA* thermometer function.¹³ However, other work by de Smit and van Duin has demonstrated that even slight weakening of the SD structure is enough to increase translation efficiency,⁵ particularly in the case of a strong SD sequence.³⁹ It is also worth noting that a previous SHAPE-based study of thermal unfolding of the D-stem within tRNA^{Asp} included the observation that during melting, the increase in the reactivity of the purine-rich side of the stem lagged behind that of the pyrimidine-rich side, potentially because of base stacking interactions between the purines.²⁸ Therefore, base stacking within the purine-rich SD sequence may contribute to its low observed SHAPE reactivity compared to that of the opposite side of the P2 stem.

In addition, further exploration of *agsA* thermometer structure with active or stalled ribosomes revealed an important mechanistic insight: ribosomes can exploit small changes in the structural ensemble to increase the level of translation of mRNAs at elevated temperatures. Specifically, we observed an increase in reactivities in the region paired to the Shine-Dalgarno site when stalled ribosomes were present, suggesting that ribosome binding can unwind the *agsA* thermometer's P2 helix to facilitate translation initiation. This happens under heat shock conditions and to a lesser degree at lower temperatures, as evidenced by the changes in reactivities surrounding the ribosome binding site at 30 °C in Figure 6c, as well as the background level of gene expression that can be detected at 30 °C. These observations reinforce the idea that the ribosome is able to take advantage of very subtle shifts in RNA structure to promote further hairpin dissolution and binding with the Shine-Dalgarno site, even when the majority of the population of RNA thermometer hairpins is well-folded.

A similar mechanism for ribosomal involvement in RNA thermometer function has also recently been suggested.⁴⁰ Narayan and colleagues' careful study of the MiniROSE RNA thermometer's structure through insertion of fluorescent 2-aminopurine residues showed only slight temperature-based structural changes in the absence of ribosomes yet a significant loss of folded structure in the presence of ribosomes at high temperatures.⁴⁰ These observations led them to propose a mechanism in which the ribosome binding promotes local melting of key regions of the MiniROSE RNA thermometer to allow for translation initiation. Moreover, a previous study of translation initiation complex formation on mRNAs containing structured RBSs showed that ribosomal protein S1 plays an important role in encouraging RNA melting to allow for docking and accommodation into the ribosomal decoding channel,⁴¹ bolstering the case for ribosome-mediated RNA unfolding.

In conclusion, we have demonstrated strong evidence that subtle differences in RNA structures mediated by temperature changes can be exploited by ribosomes to give rise to large changes in gene expression. It is interesting to speculate whether this mechanism could be important for the general control of gene expression in a temperature-dependent manner. The elegantly simple hairpin fluctuation coupled to the ribosome-mediated melting mechanism behind *agsA* thermosensing offers a potential template for the future design of synthetic RNA thermometers,^{42–44} and the combination of SHAPE-Seq with *in vitro* mechanistic studies shows how *in vivo* probing can be used to validate the structural mechanism behind regulatory RNAs.

■ ASSOCIATED CONTENT

● Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.biochem.7b01170.

Figures S1–S11 and Table S1 (PDF)

Accession Codes

RNA probing data available at the RNA Mapping Database (<https://rmdb.stanford.edu>) under structure numbers *agsAR-NA_1M7_0001*, *agsARNA_1M7_0002*, *agsAR-NA_1M7_0003*, *agsARNA_1M7_0004*, *agsAR-NA_1M7_0005*, and *agsARNA_1M7_0006*.

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Notes

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■ ABBREVIATIONS

RBS, ribosome binding site; ROSE, repression of heat shock gene expression; UTR, untranslated region; SD, Shine-Dalgarno; NMR, nuclear magnetic resonance; SHAPE-Seq, selective 2'-hydroxyl acylation analyzed by primer extension sequencing; WT, wild-type; SFGFP, Superfolder green fluorescent protein; LB, lysogeny broth; OD, optical density; 1M7, 1-methyl-7-nitroisatoic anhydride; DMSO, dimethyl sulfoxide; MFE, minimum free energy; MDS, multidimensional scaling; TIC, translation initiation complex.

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